

=> "fusion proteiin" or "fusion polypeptide"
215938 "FUSION"
8044 "FUSIONS"
220290 "FUSION"
("FUSION" OR "FUSIONS")
8 "PROTEIIN"
3 "PROTEIINS"
11 "PROTEIIN"
("PROTEIIN" OR "PROTEIINS")
0 "FUSION PROTEIIN"
("FUSION" (W) "PROTEIIN")
215938 "FUSION"
8044 "FUSIONS"
220290 "FUSION"
("FUSION" OR "FUSIONS")
90800 "POLYPEPTIDE"
52251 "POLYPEPTIDES"
123478 "POLYPEPTIDE"
("POLYPEPTIDE" OR "POLYPEPTIDES")
357 "FUSION POLYPEPTIDE"
("FUSION" (W) "POLYPEPTIDE")
L1 357 "FUSION PROTEIIN" OR "FUSION POLYPEPTIDE"

=> biotin and L1
23752 BIOTIN
99 BIOTINS
23760 BIOTIN
(BIOTIN OR BIOTINS)
L2 3 BIOTIN AND L1

=> avidine and L1
12 AVIDINE
1 AVIDINES
13 AVIDINE
(AVIDINE OR AVIDINES)
L3 0 AVIDINE AND L1

=> biotinylation and L1
1747 BIOTINYLATION
5 BIOTINYLATIONS
1749 BIOTINYLATION
(BIOTINYLATION OR BIOTINYLATIONS)
L4 0 BIOTINYLATION AND L1

=> carboxylase and L1
16353 CARBOXYLASE
1346 CARBOXYLASES
16573 CARBOXYLASE
(CARBOXYLASE OR CARBOXYLASES)
L5 1 CARBOXYLASE AND L1

=> PSTCD and L1
0 PSTCD
L6 0 PSTCD AND L1

=> " viral antigen"
120760 "VIRAL"
7 "VIRALS"
120766 "VIRAL"
("VIRAL" OR "VIRALS")
237262 "ANTIGEN"
188286 "ANTIGENS"
293896 "ANTIGEN"
("ANTIGEN" OR "ANTIGENS")

L7 3002 " VIRAL ANTIGEN"
("VIRAL" (W) "ANTIGEN")

=> surface and L7
1822535 SURFACE
357645 SURFACES
1969913 SURFACE
(SURFACE OR SURFACES)

L8 524 SURFACE AND L7

=> L1 and L8
L9 2 L1 AND L8

=> DIS L9 1- IBIB ABS

=> biotinylation
1747 BIOTINYLATION
5 BIOTINYLATIONS
L10 1749 BIOTINYLATION
(BIOTINYLATION OR BIOTINYLATIONS)

=> virus (l) antigen
283719 VIRUS
60654 VIRUSES
293949 VIRUS
(VIRUS OR VIRUSES)
237262 ANTIGEN
188286 ANTIGENS
293896 ANTIGEN
(ANTIGEN OR ANTIGENS)
L11 39847 VIRUS (L) ANTIGEN

=> L10 and L11
L12 18 L10 AND L11

L12 ANSWER 8 OF 18 CAPLUS COPYRIGHT 2003 ACS
 ACCESSION NUMBER: 1998:66044 CAPLUS
 DOCUMENT NUMBER: 128:86181
 TITLE: Labeling and selection of molecules
 INVENTOR(S): Osbourn, Jane Katharine; Derbyshire, Elaine Joy;
 McCafferty, John Gerald; Vaughan, Tristan John;
 Johnson, Kevin Stuart
 PATENT ASSIGNEE(S): Cambridge Antibody Technology Ltd., UK; Osbourn, Jane
 Katharine; Derbyshire, Elaine Joy; McCafferty, John
 Gerald; Vaughan, Tristan John; Johnson, Kevin Stuart
 SOURCE: PCT Int. Appl., 150 pp.
 CODEN: PIXXD2
 DOCUMENT TYPE: Patent
 LANGUAGE: English
 FAMILY ACC. NUM. COUNT: 1
 PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9801757	A1	19980115	WO 1997-GB1835	19970708
W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
RW: GH, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
CA 2259421	AA	19980115	CA 1997-2259421	19970708
GB 2315125	A1	19980121	GB 1997-14397	19970708
GB 2315125	B2	19980603		
AU 9734527	A1	19980202	AU 1997-34527	19970708
AU 715796	B2	20000210		
EP 906571	A1	19990407	EP 1997-930647	19970708
EP 906571	B1	20030423		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
GB 2330909	A1	19990505	GB 1999-205	19970708
GB 2330909	B2	20001227		
US 5994519	A	19991130	US 1997-889291	19970708
JP 2000517046	T2	20001219	JP 1998-504944	19970708
AT 238550	E	20030515	AT 1997-930647	19970708
US 6180336	B1	20010130	US 1998-98244	19980617
US 6342588	B1	20020129	US 1999-375314	19990816
US 2002004215	A1	20020110	US 2001-767395	20010123
US 6489123	B2	20021203		
PRIORITY APPLN. INFO.:			GB 1996-14292	A 19960708
			GB 1996-24880	A 19961129
			GB 1997-12818	A 19970618
			US 1997-889291	A3 19970708
			WO 1997-GB1835	W 19970708
			US 1998-98244	A1 19980617

AB A method of labeling mols. which includes providing in a common medium a label mol., a marker ligand able to bind a member of a specific binding pair, such as an antigen, a sbp member, an enzyme able to catalyze binding of the label mol. to other mols., the enzyme being assocd. with the marker ligand; causing or allowing binding of the marker ligand to the sbp member; and causing or allowing binding of the label mol. to other mols. in the vicinity of the marker ligand bound to the sbp member. The marker ligand may be an antibody or any specific binding mol., such as a chemokine or cytokine. A complementary member of the specific binding pair may be included, e.g. an antibody, or a diverse population of such sbp members, e.g. antibodies, may be included within

which those which bind the counterpart sbp member, e.g. **antigen**, may be labeled and subsequently isolated for manipulation and/or use. Suitable labels include biotin-tyramine with signal transfer being catalyzed by hydrogen peroxidase. Cells, **virus** particles and other moieties may be labeled, for identification or obtention of proteins which interact or are in close proximity with a particular sbp member, or of cells of interest, or for enhancement of labeling, e.g. for cell sorting.

L12 ANSWER 1 OF 18 CAPLUS COPYRIGHT 2003 ACS
ACCESSION NUMBER: 2003:102566 CAPLUS
DOCUMENT NUMBER: 138:270002
TITLE: Genetic Engineering of Streptavidin-Binding Peptide
Tagged Single-Chain Variable Fragment Antibody to
Venezuelan Equine Encephalitis Virus
AUTHOR(S): Hu, Wei-Gang; Alvi, Azhar Z.; Fulton, R. Elaine;
Suresh, Mavanur R.; Nagata, Les P.
CORPORATE SOURCE: Development Canada-Suffield, Chem. Biological Defence
Section, AB, T1A 8K6, Can.
SOURCE: Hybridoma and Hybridomics (2002), 21(6), 415-420
CODEN: HHYYBF; ISSN: 1536-8599
PUBLISHER: Mary Ann Liebert, Inc.
DOCUMENT TYPE: Journal
LANGUAGE: English
AB A recombinant gene encoding a single-chain variable fragment (scFv)
antibody against Venezuelan equine encephalitis virus (VEE) was
cloned into a prokaryotic T7 RNA polymerase-regulated expression vector.
A streptavidin-binding peptide gene fused to a 6His tag was attached
downstream to the scFv gene. The recombinant fusion protein was expressed
in bacteria as inclusion bodies that were subsequently solubilized with 8
M urea and renatured by an arginine system. Purifn. of the fusion protein
was achieved by immobilized metal affinity chromatog. ELISA and Western
blotting results revealed that the fusion protein not only retained VEE
antigen binding and specificity properties similar to those of its
parent native monoclonal antibody (MAb), but also possessed
streptavidin-binding activity. This exptl. approach can eliminate the
need for chem. biotinylation of antibodies and the risk assocd.
of antibody denaturation and can provide a stable and reproducible reagent
for rapid and efficient immunoassay of VEE when detected by horseradish
peroxidase (HRP)-conjugated streptavidin.
REFERENCE COUNT: 29 THERE ARE 29 CITED REFERENCES AVAILABLE FOR THIS
RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT